

**APPENDIX A**

Attached is a copy of an article entitled, "Thymulin and the neuroendocrine system,"  
*Peptides*, 25:139-142, 2004.



## Review

## Thymulin and the neuroendocrine system

Rodolfo G. Goya<sup>a,\*</sup>, Oscar A. Brown<sup>a</sup>, Jean-Marie Pléau<sup>b</sup>, Mireille Dardenne<sup>b</sup><sup>a</sup> Faculty of Medicine, Institute for Biochemical Research at La Plata-Histology B, National University of La Plata, La Plata, Argentina<sup>b</sup> CNRS URA 8603, Université Paris V, Hôpital Necker, Paris, France

Received 21 July 2003; received in revised form 3 November 2003; accepted 4 November 2003

**Abstract**

Thymulin is a thymic hormone exclusively produced by the thymic epithelial cells. It consists of a nonapeptide component coupled to the ion zinc, which confers biological activity to this molecule. After its discovery in the early 1970, thymulin was characterized as a thymic hormone involved in several aspects of intra- and extrathymic T-cell differentiation. Subsequently, it was demonstrated that thymulin production and secretion is strongly influenced by the neuroendocrine system. Conversely, an emerging core of information points to thymulin as a hypophysiotropic peptide. Here we review the evidence supporting the hypothesis that thymulin is an important player in the hypophyso-thymic axis.

© 2003 Elsevier Inc. All rights reserved.

**Keywords:** Thymulin; Hypophysiotropic activities; Nude mouse; Aging; Dyshomeostasis; Neuroendocrine imbalances; Gene therapy

**1. Introduction**

It is now well-established that the immune system is functionally linked to the nervous and endocrine systems thus constituting an integrated homeostatic network [23]. Within this network, the neuroendocrine system monitors and controls the physical and chemical variables of the internal milieu. On its part, the immune system perceives, through antigenic recognition, an internal image of the macromolecular and cellular components of the body and reacts to alterations of this image, effectively participating of the “biological” homeostasis of the organism. The relevance of the thymus in this network becomes evident when one observes the immune and neuroendocrine consequences of neonatal thymectomy or congenital absence of the thymus in certain animal species (see below). In this context, the existence of a neuroendocrine-thymic axis is well-documented (for a review, see [47]).

Thymulin is a thymic hormone involved in several aspects of intra- and extrathymic T-cell differentiation [1]. Thymulin, which is exclusively produced by the thymic epithelial cells (TEC), consists of a biologically inactive nonapeptide component (*facteur thymique sérique* or FTS) coupled in an equimolecular ratio to the ion zinc [20], which confers biological activity to this molecule [13]. In the present

paper, we will review the evidence indicating that thymulin is an important player in the hypophyso-thymic axis.

**2. Evidence for a pituitary-thymulin axis**

The control of thymulin secretion seems to be dependent on a complex network of events. Initial studies showed that the hormone itself exerts a controlling feedback effect on its own secretion both *in vivo* and *in vitro* [9,48].

Additionally, thymulin production and secretion is influenced directly or indirectly by the neuroendocrine system. For instance, growth hormone (GH) can influence thymulin synthesis and secretion. *In vitro*, hGH can stimulate thymulin release from TEC lines [50] which are known to possess specific receptors for GH [2]. Animal studies have shown that treatment of aged dogs with bovine GH partially restored their low thymulin serum levels [21]. In old mice, treatment with ovine GH increased their low circulating thymulin and enhanced the concanavalin A (Con A)-dependent proliferative response of thymocytes, as well as interleukin-6 production [26]. In old rats, combined treatment with GH and thyroxine (T<sub>4</sub>) was also able to restore partially their reduced thymulin levels [27]. In clinical studies, it was reported that in congenitally GH-deficient children, who consistently exhibited low plasma thymulin levels, GH therapy succeeded in increasing thymic hormone levels to near normal values [41]. Acromegalic middle aged patients have elevated thymulin serum levels compared to age-matched normal sub-

\* Corresponding author. Tel.: +54-221-425-6735;  
fax: +54-221-425-0924.

E-mail address: rgoya@netwerk.com.ar (R.G. Goya).

jects [41,50]. It is likely that these effects of GH are mediated, at least in part, by insulin-like growth factor 1 (IGF-1) as suggested by the fact that the GH-induced enhancement of thymulin production *in vitro* could be prevented by previous treatment with antibodies against IGF-1 or IGF-1 receptor [50].

There is also evidence for a prolactin (PRL)-thymulin axis. Thus, it is known that TEC possess PRL receptors [12] and that PRL can stimulate thymulin synthesis and secretion both *in vitro* and *in vivo* [15]. Furthermore, administration of PRL to old mice elevated their reduced circulating levels of thymulin [15].

The thyroid axis also influences thymulin secretion. Thus, T<sub>4</sub> has been shown to stimulate thymulin synthesis and secretion in mice [17]. *In vivo* treatment of mice with triiodothyronine (T<sub>3</sub>) enhanced thymulin secretion whereas treatment of the animals with propylthiouracil, an inhibitor of thyroid hormone synthesis, decreased their circulating thymulin levels [49]. In humans, hyperthyroidism brings about an increase in circulating thymulin levels whereas hypothyroid patients show depressed levels of this thymic hormone [18]. In *in vitro* studies, it was shown that thyroid hormones stimulate thymulin secretion by a direct action on TEC [40,51]. Interestingly, it has been shown that treatment of aged animals with T<sub>4</sub> can reverse their decreased thymulin levels [17,40].

Although there are no studies documenting a direct effect of gonadotropins or adrenocorticotrophic hormone (ACTH) on thymulin secretion, gonadectomy or adrenalectomy in mice are known to induce a transient decrease in serum thymulin levels. This effect is potentiated by the simultaneous removal of the adrenals and gonads [14]. In TEC cultures, it was shown that exposure to physiological levels of glucocorticoids or gonadal steroids enhanced thymulin concentration in the cell supernatants [46].

Although there is no rigorous evidence proving the existence of hypothalamic factors able to influence thymulin production by a direct action on TEC, there are two studies which suggest that this may be the case. Treatment of old mice with hypothalamic extracts from young mice resulted in reappearance of detectable levels of circulating thymulin [19]. Hypothalamic and pituitary extracts from young mice stimulated thymulin release from TEC cultures but this stimulation declined when the pituitary and hypothalamic extracts were obtained from old mice [28].

### 3. Hypophysiotropic activity of thymulin

The multilateral influence that the neuroendocrine system exerts on thymulin secretion suggests that this metallopeptide could in turn be part of a feedback loop acting on neuroendocrine structures. This possibility is now supported by a significant body of evidence indicating that thymulin possesses hypophysiotropic activity. Thus, thymulin has been shown to stimulate luteinizing hormone (LH) re-

lease from perfused rat pituitaries [53] and ACTH from incubated rat pituitary fragments, the latter being an effect mediated by intracellular cyclic adenosine monophosphate (cAMP) and cyclic guanosine monophosphate (cGMP) accumulation [31]. Thymulin has been found to stimulate GH, PRL, thyrotropin (TSH) and gonadotropin release in dispersed rat pituitary cells at doses from 10<sup>-8</sup> to 10<sup>-3</sup> M [4–6] whereas others have reported that thymulin doses of 10<sup>-11</sup> M stimulate LH, inhibit PRL release and have no effect on GH secretion in incubated rat pituitary fragments [31]. The stimulatory effect of thymulin on hormone release in rat pituitary cells declines with the age of the cell donor which suggests that aging brings about a desensitization of the pituitary gland to thymic signals [4–6].

Interestingly, congenitally athymic (nude) mice display a number of neuroendocrine deficiencies consistent with their lack of circulating thymulin (i.e., while thymulin generally stimulates pituitary hormone secretion, athymia seems to be associated with a functional panhypopituitarism). Thus, in a series of studies it was demonstrated that in nude CD-1 male mice, TSH, PRL, GH and gonadotropin responses to immobilization and cold stress are reduced as also are serum basal levels of the same hormones [25,29,30]. Nude female mice show significantly reduced levels of circulating and pituitary gonadotropins, a fact that seems to be causally related to a number of reproductive derangements described in these mutants [43]. Thus, in homozygous (nu/nu) females the times of vaginal opening and first ovulation are delayed [3], fertility is reduced [43], and follicular atresia is increased such that premature ovarian failure results [37]. Similar abnormalities result from neonatal thymectomy of normal female mice [39,42]. There is *in vitro* and *in vivo* evidence suggesting that thymulin plays a role in the regulation of female spontaneous puberty, possibly through its effects on ovarian steroidogenesis [32]. Thymulin also modulates gonadotropin-induced testicular steroidogenesis [52].

A functional impairment of the hypothalamo-adrenal axis has been reported in nude mice suggesting that humoral thymic factors may play a role in the physiology of this axis [11].

Since thymulin has been shown to be active on the nervous system (for a review, see [45]) and hypothalamic factors appear to influence thymulin secretion (see above), the peptide might exert a direct or indirect feedback effect at hypothalamic level.

### 4. The prospect of gene therapy for thymulin in thymus-depressing pathologies

Studies in animals have demonstrated that aging brings about a severe involution of the thymus and that circulating levels of thymulin are very low in old animals [21,26,27]. In healthy humans, serum thymulin levels remain high until 10–15 years of age, then fall progressively until 35–40 years of age, remaining at very low plateau levels afterwards

[10,16,35]. There is also a number of clinical situations associated with markedly low levels of circulating thymulin. These situations include, but are not limited to, AIDS [34], Di George syndrome [35] and other immunodeficiencies [33,35] as well as Down's syndrome [16].

As thymus involution is very difficult to reverse by pharmacological means, the prospect of implementing thymic hormone gene therapy appears as an interesting avenue of research aimed at restoring circulating levels of thymic hormones when thymus function is compromised [24]. Unfortunately, up to date the gene coding for thymulin has not been cloned, a situation that hinders the application of gene therapy for this thymic hormone. A possible way to overcome this problem is to construct an "artificial gene" coding for thymulin. Previous studies demonstrated that this can be done in bacteria. Thus, a synthetic DNA sequence coding for thymulin was inserted into a bacterial expression vector and successfully used to obtain large quantities of purified thymulin retaining full biological activity [8]. Consequently, an adenoviral vector (adenoviral vectors are highly efficient gene delivery systems) harboring this synthetic gene for thymulin could be constructed. As the synthetic gene product must be targeted to the secretory pathway, so that thymulin may be secreted by the transduced cells, a secretory signal DNA sequence would have to be attached upstream the synthetic gene. This type of strategy has been successfully implemented for other proteins [36].

Skeletal muscle has been shown to be a well-suited tissue for efficient viral-vector-mediated peptide hormone gene transfer as well as for long-term regulated expression and secretion of the transgenic peptide [38,44]. Therefore, the adenoviral vector carrying the thymulin-encoding DNA sequence could be injected intramuscularly in an appropriate animal model as the nude mouse. Transduced myocytes should then begin to act as an ectopic source of thymulin thus restoring circulating thymulin levels. Additionally, the thymulin synthetic gene carried by the adenoviral vector could be placed under the control of a regulable promoter. That is, a promoter that can be turned on or off by the administration of small molecules like the antibiotic tetracycline or the steroid mifepristone, both of which have been successfully used for this purpose [7,22]. The use of a regulable vector system for the thymulin synthetic gene would allow to control the circulating levels of the hormone, lowering or increasing them as physiological or pathological circumstances demand.

## 5. Concluding remarks

While the existence of a neuroendocrine-thymic axis is well-established, the evidence reviewed here strongly suggests that thymulin plays a physiological role as part of an ascending feedback loop in this axis. Consequently, it would be reasonable to expect that pathological situations associated with thymulin deficiency (or overproduction) may alter

the neuroendocrine balance. Within this context, gene therapy for thymulin may succeed in restoring some of the immune, endocrine, and reproductive abnormalities associated with thymus deficiency.

## Acknowledgments

The authors are grateful to Ms. Yolanda Sosa for secretarial assistance with the manuscript. Part of our own work reported here was aided by Grant PICT98-4594 to R.G.G. and Grant Antorchas to O.A.B. R.G.G. and O.A.B. are permanent scientists of the Argentine Research Council (CONICET).

## References

- [1] Bach JF. Thymulin (FTS-Zn). *Clin Immunol Allergy* 1983;3:133–56.
- [2] Ban E, Gagnerault MC, Jammes H, Postel-Vinay MC, Haour F, Dardenne M. Specific binding sites for growth hormone in cultured mouse thymic epithelial cells. *Life Sci* 1991;48:2141–8.
- [3] Besedovsky HO, Sorkin E. Thymus involvement in female sexual maturation. *Nature* 1974;249:356–8.
- [4] Brown OA, Sosa YE, Bolognani F, Goya RG. Thymulin stimulates prolactin and thyrotropin release in an age-related manner. *Mech Age Dev* 1998;104:249–62.
- [5] Brown OA, Sosa YE, Dardenne M, Pléau JM, Goya RG. Growth hormone-releasing activity of thymulin: effects of age. *Neuroendocrinology* 1999;69:20–7.
- [6] Brown OA, Sosa YE, Dardenne M, Pléau JM, Goya RG. Studies on the gonadotropin-releasing activity of thymulin: changes with age. *J Gerontol (Biol Sci)* 2000;55:B170–176.
- [7] Burcin MM, Schiedner G, Kochanek S, Tsai SY, O'Malley BW. Adenovirus-mediated regulable target gene expression in vivo. *Proc Natl Acad Sci USA* 1999;96:355–60.
- [8] Calenda A, Cordonnier A, Lederer F, Le DKH, Pléau JM. Production of biologically active thymulin in *Escherichia coli* through expression of a chemically synthesized gene. *Biotechnol Lett* 1988;10:155–60.
- [9] Cohen S, Berrih S, Dardenne M, Bach JF. Feed-back regulation of the secretion of a thymic hormone (thymulin) by human thymic epithelial cells in culture. *Thymus* 1986;8:109–19.
- [10] Consolini R, Legitimo A, Calleri A, Milani M. Distribution of age-related thymulin titres in normal subjects through the course of life. *Clin Exp Immunol* 2000;121:444–7.
- [11] Daneva T, Spinedi E, Hadid R, Gaillard RC. Impaired hypothalamo-pituitary-adrenal axis function in swiss nude athymic mice. *Neuroendocrinology* 1995;62:79–86.
- [12] Dardenne M, Kelly PA, Bach JF, Savino W. Identification and functional activity of Prl receptors in thymic epithelial cells. *Proc Natl Acad Sci USA* 1991;88:9700–4.
- [13] Dardenne M, Pleau JM, Nabarra B, Lefrancier P, Derrien M, Choay J, et al. Contribution of zinc and other metals to the biological activity of serum thymic factor (FTS). *Proc Natl Acad Sci USA* 1982;79:5370–3.
- [14] Dardenne M, Savino W, Duval D, Kaiserlian D, Hassid J, Bach JF. Thymic hormone-containing cells. VII. Adrenals and gonads control the in vivo secretion of thymulin and its plasmatic inhibitor. *J Immunol* 1986;136:1303–8.
- [15] Dardenne M, Savino W, Gagnerault MC, Itoh T, Bach JF. Neuroendocrine control of thymic hormonal production. I. Prolactin stimulates in vivo and in vitro the production of thymulin by human and murine thymic epithelial cells. *Endocrinology* 1989;125:3–12.

[16] Fabris N, Amadio L, Licastro F, Mocchegiani E, Zannotti M, Franceschi C. Thymic hormone deficiency in normal ageing and Down's syndrome: is there a primary failure of the thymus? *Lancet* 1984;1:983–6.

[17] Fabris N, Mocchegiani E. Endocrine control of thymic serum factor production in young-adult and old mice. *Cell Immunol* 1985;91:325–35.

[18] Fabris N, Mocchegiani E, Mariotti S, Pacini F, Pinchera A. Thyroid function modulates thymic endocrine activity. *J Clin Endocrinol Metab* 1986;62:474–8.

[19] Folch H, Eller G, Mena M, Esquivel P. Neuroendocrine regulation of thymus hormones: hypothalamic dependence of FTS level. *Cell Immunol* 1986;102:211–6.

[20] Gastinel LN, Dardenne M, Pléau JM, Bach JF. Studies on the zinc-binding site to the serum thymic factor. *Biochim Biophys Acta* 1984;797:147–55.

[21] Goff BL, Roth JA, Arp LH, Incefy GS. Growth hormone treatment stimulates thymulin production in aged dogs. *Clin Exp Immunol* 1987;68:580–7.

[22] Gossen M, Bujard H. Tight control of gene expression in mammalian cells by tetracycline responsive promoters. *Proc Natl Acad Sci USA* 1992;88:5547–51.

[23] Goya RG. The immune-neuroendocrine homeostatic network and ageing. *Gerontology* 1991;37:208–13.

[24] Goya RG, Cónsole GM, Hereñú CB, Brown OA, Rimoldi OJ. Thymus and aging: potential of gene therapy for restoration of endocrine thymic function in thymus-deficient animal models. *Gerontology* 2002;48:325–8.

[25] Goya RG, Cónsole GM, Sosa YE, Gómez Dumm CLA, Dardenne M. Altered functional responses with preserved morphology of gonadotrophic cells in congenitally athymic mice. *Brain Behav Immun* 2001;15:85–92.

[26] Goya RG, Gagnerault MC, Leite de Moraes MC, Savino W, Dardenne M. In vivo effects of growth hormone on thymus function in aging mice. *Brain Behav Immun* 1992;6:341–54.

[27] Goya RG, Gagnerault MC, Sosa YE, Bevilacqua JA, Dardenne M. Effects of growth hormone and thyroxine on thymulin secretion in aging rats. *Neuroendocrinology* 1993;58:338–43.

[28] Goya RG, Gagnerault MC, Sosa YE, Dardenne M. Reduced ability of pituitary extracts from old mice to stimulate thymulin secretion in vitro. *Mech Age Dev* 1995;83:143–54.

[29] Goya RG, Sosa YE, Cónsole GM, Dardenne M. Altered thyrotropic and somatotropic responses to environmental challenges in congenitally athymic mice. *Brain Behav Immun* 1995;9:79–86.

[30] Goya RG, Sosa YE, Cónsole GM, Dardenne M. Altered regulation of serum prolactin in nude mice. *Med Sci Res* 1996;24:279–80.

[31] Hadley AJ, Rantle CM, Buckingham JC. Thymulin stimulates corticotrophin release and cyclic nucleotide formation in the rat anterior pituitary gland. *Neuroimmunomodulation* 1997;4:62–9.

[32] Hinojosa L, Chavira R, Dominguez R, Rosas P. Effects of thymulin on spontaneous puberty and gonadotrophin-induced ovulation in prepubertal normal and hypothymic mice. *J Endocrinol* 1999;163:255–60.

[33] Incefy GS, Dardenne M, Pahwa S, Grimes E, Pahwa RN, Smithwick E, et al. Thymic activity in severe combined immuno-deficiency diseases. *Proc Natl Acad Sci USA* 1977;74(3):1250–3.

[34] Incefy GS, Pahwa S, Pahwa R, Sarngadharan MG, Menez R, Fikrig S. Low circulating thymulin-like activity in children with AIDS and AIDS-related complex. *AIDS Res* 1986;2(2):109–16.

[35] Iwata T, Incefy GS, Cunningham-Rundles S, Cunningham-Rundles C, Smithwick E, Geller N, et al. Circulating thymic hormone activity in patients with primary and secondary immunodeficiency diseases. *Am J Med* 1981;71:385–94.

[36] Kaether C, Gerdes HH. Visualization of protein transport along the secretory pathway using green fluorescent protein. *FEBS Lett* 1995;369:267–71.

[37] Lintern-Moore S, Pantelouris EM. Ovarian development in athymic nude mice. I. The size and composition of the follicle population. *Mech Age Dev* 1975;4:385–90.

[38] Marshall DJ, Leiden JM. Recent advances in skeletal-muscle-based gene therapy. *Curr Opin Genet Dev* 1998;8:360–5.

[39] Michael SD, Taguchi O, Nishizuka Y. Effects of neonatal thymectomy on ovarian development and plasma LH, FSH, GH and PRL in the mouse. *Biol Reprod* 1980;22:343–50.

[40] Mocchegiani E, Amadio L, Fabris N. Neuroendocrine-thymus interactions. I. In vitro modulation of thymic factor secretion by thyroid hormones. *J Endocrinol Invest* 1990;13:139–47.

[41] Mocchegiani E, Paolucci P, Balsamo A, Cacciari E, Fabris N. Influence of growth hormone on thymic endocrine activity in humans. *Horm Res* 1990;33:7–14.

[42] Nishizuka Y, Sakakura T. Ovarian dysgenesis induced by neonatal thymectomy in the mouse. *Endocrinology* 1971;89:889–93.

[43] Rebar RW, Morandini IC, Erickson GF, Petze JE. The hormonal basis of reproductive defects in athymic mice. *Endocrinology* 1981;108:120–6.

[44] Rivera VM, Ye X, Courage NL, Sachar J, Cerasoli Jr F, Wilson JM, et al. Long-term regulated expression of growth hormone in mice after intramuscular gene transfer. *Proc Natl Acad Sci USA* 1999;96:8657–62.

[45] Safieh-Garabedian B, Kanaan SA, Jabbur SJ, Saade NE. Cytokine-mediated or direct effects of thymulin on the nervous system as assessed by pain-related behavior. *Neuroimmunomodulation* 1999;6:39–44.

[46] Savino W, Bartoccioni E, Homo-Delarche F, Gagnerault MC, Itoh T, Dardenne M. Thymic hormone containing cells. IX. Steroids in vitro modulate thymulin secretion by human and murine thymic epithelial cells. *J Steroid Biochem* 1988;30:479–84.

[47] Savino W, Dardenne M. Neuroendocrine control of thymus physiology. *Endocr Rev* 2000;21:412–43.

[48] Savino W, Dardenne M, Bach JF. Thymic hormone containing cells. III. Evidence for a feed-back regulation of the secretion of the serum thymic factor (FTS) by thymic epithelial cells. *Clin Exp Immunol* 1983;52:7–12.

[49] Savino W, Wolf B, Aratan-Spire S, Dardenne M. Thymic hormone containing cells. IV. Fluctuations in the thyroid hormone levels in vivo can modulate the secretion of thymulin by the epithelial cells of young mouse thymus. *Clin Exp Immunol* 1984;55:629–35.

[50] Timsit J, Savino W, Safieh B, Chanson P, Gagnerault MC, Bach JF, et al. GH and IGF-I stimulate hormonal function and proliferation of thymic epithelial cells. *J Clin Endocrinol Metab* 1992;75:183–8.

[51] Villa-Verde DMS, Mello-Coelho V, Farias de Oliveira DA, Dardenne M, Savino W. Pleiotropic influence of triiodothyronine on thymus physiology. *Endocrinology* 1993;133:867–75.

[52] Wise T. In vitro and in vivo effects of thymulin on rat testicular steroid synthesis. *J Steroid Biochem Mol Biol* 1998;66:129–35.

[53] Zaidi SA, Kendall MD, Gillham B, Jones MT. The release of LH from pituitaries perfused with thymic extracts. *Thymus* 1988;12:253–64.